Dual Roles for Immunity in Gastrointestinal Cancers

Cristina Ferrone and Glenn Dranoff

ABSTRACT

Histopathologic examination reveals that most human tumors are associated with diverse immune cell infiltrates, but the roles of host reactions in disease pathogenesis and prognosis remain to be fully clarified. Recent investigations in genetically engineered murine tumor models have uncovered dual functions for immune responses during cancer development and progression. Alterations in tumor cell gene expression profiles and coding sequences may trigger the activation of cytotoxic lymphocytes, which act to restrain tumor growth. In contrast, persistent inflammatory reactions, which may be driven by infection, environmental toxins, or impaired immune regulation, create a microenvironment that fosters tumor cell growth, survival, invasion, and dissemination. The dynamic interplay of these competing responses appears to be a critical event in cancer pathogenesis, with tumor promotion and immune evasion proving dominant in clinically evident disease. Nonetheless, longitudinal studies of patient cohorts have demonstrated that particular histopathologic and genetic signatures of cytotoxic lymphocyte reactions provide important prognostic information. Here, we discuss the dual roles of immunity in cancer development, focusing on gastrointestinal malignancies, given the depth of recent insights into the mechanisms underlying these tumors.

J Clin Oncol 28:4045-4051. © 2010 by American Society of Clinical Oncology

From the Massachusetts General Hospital; Cancer Vaccine Center and Dana-Farber Cancer Institute; Brigham and Women's Hospital; and Harvard

Submitted January 4, 2010; accepted May 25, 2010; published online ahead of print at www.jco.org on July 19, 2010

Medical School, Boston, MA.

Authors' disclosures of potential conflicts of interest and author contributions are found at the end of this article.

Corresponding author: Glenn Dranoff, MD, Dana-Farber Cancer Institute, 44 Binney St, Dana 520C, Boston, MA 02115; e-mail: glenn_dranoff@dfci .harvard.edu.

© 2010 by American Society of Clinical Oncology

0732-183X/10/2826-4045/\$20.00 DOI: 10.1200/JCO.2010.27.9992

INTRODUCTION

Virchow's perceptive observation of the close association between cancer and inflammation in the nineteenth century anticipated the current interest in the role of immunity in tumor pathogenesis. Recent insights into the dynamics of the tumor microenvironment have begun to clarify the mechanisms underlying tumor-promoting inflammation, which bears striking similarities, as Harold Dvorak² articulated, to wounds that fail to heal. Approximately 20% of cancer deaths worldwide are currently linked to unresolved infection and/or inflammation, with gastrointestinal malignancies constituting a significant proportion of this disease burden.¹ Among the most frequent associations are colorectal carcinoma and inflammatory bowel disease, gastric carcinoma and Helicobacter pylori infection, hepatocellular carcinoma and chronic hepatitis of diverse etiologies, pancreatic carcinoma and chronic pancreatitis, esophageal carcinoma and chronic gastric reflux, anal carcinoma and human papilloma virus infection, and cholangiocarcinoma and sclerosing cholangitis.3-9

Notwithstanding these examples of aberrant immunity fostering tumor initiation and evolution, substantial data also support a protective role for immunity in tumor suppression. Paul Ehrlich's¹⁰ original notion of a cancer immunosurveillance sys-

tem at the turn of the twentieth century has been refined, through elegant studies of cancer development in genetically engineered immune deficient mice, to the contemporary, broader concept of immunoediting. In this scheme, host immunity may eliminate incipient tumors or restrain the growth of established tumors through effectuating a state of equilibrium; disease progression reflects escape from immune control. Several recent clinicopathologic studies provide suggestive evidence that immunoediting might be operative in diverse human malignancies as well.

Here, we highlight progress in elucidating these dual roles of immunity in cancer pathogenesis, with a focus on selected gastrointestinal cancers. Colorectal carcinoma, hepatocellular carcinoma, and gastric carcinoma have been chosen for detailed discussion since progress in understanding these tumors is proceeding at an especially rapid pace. Moreover, the principles revealed with these tumors are likely to be operative in other cancers as well. We first will examine the mechanisms underlying tumor-promoting inflammation, and then turn to the tumor protective pathways, which are tightly linked to patient outcomes. A deeper understanding of host-tumor interactions should advance the crafting of new therapies aimed at more effectively interfering with tumor-promoting inflammation or intensifying immune-mediated tumor

destruction. Such strategies might additionally prove complementary to targeted treatments directed at defects intrinsic to cancer cells.

MECHANISMS OF TUMOR-PROMOTING INFLAMMATION

Overview

Unresolved inflammation generates a microenvironment that is favorable for cellular transformation and the propagation of invasive disease. 1,11 Chronic tissue damage triggers a repair response that includes the production of growth and survival factors, tissue remodeling enzymes, angiogenic cytokines, and immune regulatory networks. The release of inflammatory cell-derived reactive oxygen species coupled with stimulated epithelial cell proliferation creates an elevated risk of mutagenesis. Persistently stressed cells may undergo selection for rare variants that manifest increased survival within the inflammatory milieu. The induction of an epithelial-to-mesenchymal transition as part of a program for wound healing may also favor the acquisition of stem cell-like features and invasive capabilities. These properties, together with new blood vessel formation elicited by injury, may foster the evolution of metastasis. Cross-talk between neoplastic cells and immune elements throughout the smoldering inflammation perpetuates the transforming environment, which provides the evolving tumor cells with sufficient opportunity to acquire mutations and epigenetic alterations that are necessary for cell autonomy. Below, we illustrate these complex dynamics within the context of specific gastrointestinal tumors.

Colorectal Cancer

The pathogenesis of inflammatory bowel disease (IBD) is multifactorial and represents a complicated interplay among immune components that predispose to invasive cancer. Patients with IBD, especially those with ulcerative colitis, have a significantly increased risk of colorectal cancer, with an incidence approaching 43% in the 25 to 35 years from the time of diagnosis.³ Blockade of tumor necrosis factor (TNF) - α with antibodies or soluble receptors is an important component of IBD therapy, and despite its name, TNF- α appears to function as a potent tumor promoter. TNF- α enhances cancer cell survival and augments vascular permeability, facilitating extravasation of tumor cells into the circulation and systemic dissemination.¹² In mouse models, the administration of dextran sulfate sodium (DSS) followed by azoxymethane (AOM) recapitulates important features of human IBD, including the progression to colon cancer. ^{13,14} This regimen elicits increased intracolonic TNF- α expression, with infiltration of inflammatory cells harboring the TNF- α receptor p55. ¹⁵ TNF- α receptor p55–deficient mice treated with DSS and AOM develop less inflammation in the colon and form fewer colonic tumors compared to wild-type animals. Furthermore, the infusion of a TNF- α antagonist to DSS and AOM treated wild-type mice reduces the number of infiltrating inflammatory cells, particularly neutrophils, mast cells, and macrophages, with a concomitant reduction in carcinoma formation. These findings highlight a key role for TNF- α in chemically induced colitis and carcinogenesis.

One of the major signaling events that TNF- α triggers in both tumor and immune cells is the nuclear factor- κ B (NF- κ B) pathway. NF- κ B refers to a family of transcription factors that regulate multiple inflammatory cytokines, adhesion molecules, antiapoptotic proteins, angiogenic factors, and the prostaglandin biosynthetic pathway

(through cyclo-oxygenase-2 [COX-2]).16 NF-κB proteins are normally retained in the cytoplasm through a set of inhibitory proteins, but following receptor-mediated activation of IkB kinases (IKK), the inhibitors are degraded, thereby allowing nuclear import of the transcription factors. ¹⁷ Studies of mice deficient in IKK- β have revealed a critical role for this pathway in AOM/DSS-induced colon tumors. 18 Selective deletion of the IKK-B gene in intestinal epithelial cells reduces tumor formation, in part because of an increase in enterocyte apoptosis due to the lack of Bcl-2 and Bcl-xL. IKK-β deletion in myeloid cells also inhibits tumor progression, which reflects the diminished production of inflammatory mediators, such as interleukin (IL) -6 and epithelial growth factors. These results are consistent with clinical data indicating that nonsteroidal anti-inflammatory drugs, which antagonize the NF-κB target gene COX-2, reduce the risk of colorectal carcinoma by 75% to 81% in patients with IBD. 19,20 Furthermore, the Cancer Prevention Study II Nutrition Cohort, which included more than 18,000 patients, documented a 30% decrease in sporadic colon cancer incidence with the use of a daily adult strength aspirin for over 5 years.²¹ Together, these investigations reveal a dual requirement for NF-kB signaling in both the target cell and the inflammatory cells during the pathogenesis of colon cancer.

In addition to TNF- α , NF- κ B signaling may be activated during colon carcinogenesis through toll-like receptors (TLRs) that are expressed on various myeloid cells, some lymphocytes, and intestinal epithelia.²² TLRs may be stimulated by conserved biochemical structures expressed by commensal flora. Most TLR signaling proceeds through the adaptor protein MvD88, which is required for the subsequent activation of NF-kB. When mice deficient in MyD88 were introgressed with multiple intestinal neoplasia mice that harbor mutations in the Apc tumor suppressor, fewer polyps were observed compared to mice with intact MyD88 function. 23-26 The major contribution of MyD88 was on polyp growth and progression rather than tumor initiation, reminiscent of the impact of myeloid-specific deletion of IKK-B mentioned earlier. Expression profiling analysis revealed that MyD88-dependent signaling contributed to the induction of both tumor-specific modifier genes and genes involved in intestinal tissue repair. These results highlight the close link between wound responses and tumor formation.

A second pathway that functions in concert with NF-κB involves the signal transducer and activator of transcription proteins (STATs). These factors relay signals from extracellular stimuli and act as transcription factors that regulate many genes involved in tumor progression, including cell cycle proteins, antiapoptotic molecules, angiogenic factors, and matrix metalloproteinases. 27-30 There are seven known members of the STAT family, but STAT-3 in particular has been linked to tumor-promoting inflammation. STAT-3 may become activated in tumor cells initially in response to a variety of growth factors, such as IL-6, IL-10, and epidermal growth factor. However, once activated, STAT-3 triggers the production of additional soluble factors that create a feed-forward loop that involves both tumor cells and stromal components. This circuitry creates a procarcinogenic microenvironment that is maintained throughout disease progression. ²⁸⁻³⁰ Interestingly, an enterotoxigenic strain of *Bacteroides* fragilis that was recently implicated in colon carcinogenesis in both humans and mice is a potent inducer of STAT-3 activation.³¹ The critical role of this transcription factor in colon carcinogenesis is also underscored by the reduced incidence of AOM/DSS triggered tumors in STAT-3 deficient mice. 13,14,30

STAT-3 signaling has a decisive influence on the mixture of cytokines present in the tumor microenvironment, favoring the production of tumor-promoting factors. IL-6 is one of these critical mediators, as tumor growth can be attenuated in murine models through the pharmacologic administration of an IL-6 inhibitor.³⁰ Since IL-6 serum levels correlate with tumor size in patients with colon cancer, 32-34 these findings have potential clinical relevance. A second important STAT-3 associated cytokine is IL-23, which is expressed at high levels in human colon carcinoma tissues.³⁵ IL-23 enhances macrophage and granulocyte infiltration that perpetuates inflammation, and cooperates with IL-6 and TGF- β for the differentiation of a newly identified CD4⁺ T cell subset called Th17 cells. These lymphocytes produce high levels of the cytokines IL-17A and IL-17F that promote angiogenesis, whereas mice lacking these factors show reduced tumor formation. 36,37 IL-23 also limits the development of protective antitumor responses through inhibiting the recruitment of cytotoxic CD8⁺ T lymphocytes and instead favoring forkhead box protein P3 (FoxP3⁺) regulatory T cells (Tregs) that further antagonize antitumor cytotoxicity. 38 Through this complex array of soluble factors and cellular subsets, STAT-3 is a major driver of tumor promoting inflammation, and the development of small molecules that inhibit STAT-3 function might afford significant clinical benefits.

In addition to excessive production of inflammatory mediators, IBD can also result from a deficiency in negative immune regulators. Recent genetic analysis uncovered loss of function mutations in the receptor for IL-10 in some families with early onset IBD.³⁹ These results are in accordance with work in murine models that established a requirement for IL-10 in intestinal immune homeostasis; indeed, mice deficient in this cytokine or receptor develop severe enterocolitis that progresses to colon carcinoma. 40-42 IL-10 has pleiotropic effects on lymphocytes, granulocytes, and macrophages, which together result in a dampening of inflammatory responses. 43,44 Excessive intestinal inflammation and colon cancer similarly occur in mice that are deficient for TGF- β , whereas mutations in the TGF- β receptor are frequently detected in patients. 32,45 Intestinal inflammation may also reflect a deficiency in Tregs, a specialized CD4⁺ T cell subset expressing the FoxP3 transcription factor, which normally function to restrain innate and adaptive immune reactions. Indeed, the adoptive transfer of Tregs can mediate therapeutic effects in some mouse models, including multiple intestinal neoplasia mice with mutations in the Apc-tumor suppressor gene. 46,47 The reversal of polyposis in this model was associated with a decrease in COX-2 expression, which is consistent with the therapeutic effects of COX-2 inhibitors in humans, as well as a reduction in mastocytosis. 48-50

Gastric Cancer

Gastric cancer is the second most common cancer worldwide, and chronic inflammation appears to drive the progression from chronic gastritis to gastric atrophy, intestinal metaplasia, dysplasia, and ultimately gastric cancer. While infection with *Helicobacter pylori* is very common in human populations, only approximately 1% of exposed individuals develop gastric cancer in response to persistent infection. The subset of patients who progress to gastric cancer appear to have polymorphisms in proinflammatory cytokines, particularly IL-1 β , that leads to enhanced levels after bacterial infection, although pathogen specific factors also play important roles. IL-1 β has profound effects on inflammation and immunity, in part by in-

ducing MyD88-dependent NF- κ B activation, which may engender myeloid-derived suppressor cells that promote tumor angiogenesis and restrain protective cytotoxic lymphocyte reactions. ^{54,55} In a transgenic mouse model of gastric cancer, the administration of an IL-1 receptor antagonist blocked the progression from chronic gastritis to invasive cancer, in conjunction with the inhibition of myeloid-derived suppressor cell mobilization and recruitment. ⁵⁶

Helicobacter-associated gastric tumors show some common pathogenetic mechanisms as the colitis-induced colon tumors. Indeed, bacterial infection activates STAT-3 signaling to promote stomach carcinomas, ⁵⁷ whereas IL-10 deficient mice develop a more severe form of gastritis. ⁵⁸⁻⁶⁰ In this system, CD4⁺ T cells and interferon gamma cooperate to trigger severe inflammation. ⁶¹

Hepatocellular Carcinoma

Chronic hepatitis due to infection with hepatitis B and C viruses, alcohol, or toxins is the major risk factor for the development of hepatocellular carcinoma. One informative model for investigating the underlying mechanisms is the Mdr2-knockout mouse, which manifests progression from periductular and periportal inflammatory infiltrates to dysplasia, dysplastic nodules, invasive carcinoma, and metastasis. 62 NF- κ B activation, as measured by p65 nuclear immunostaining, is observed in this system, and this is blocked with the administration of either an anti-TNF- α antibody or COX-2 inhibitors. As in the colitis model, TNF- α appears to promote tumor formation through NF-κB mediated survival pathways in hepatocytes. However, in contrast to these findings, mice lacking IKK- β selectively in hepatocytes showed a marked increase in susceptibility to diethylnitrosamine-induced liver cancers. 63 Tumor formation in this model reflects increased hepatocyte proliferation in response to damage, with Jun kinase signaling and IL-6 production contributing to cell growth. Treatment of mice with antioxidants attenuated tumor development, in part through inhibiting hepatocyte turnover. 63

While the specific role of NF- κ B in this system remains to be clarified fully, other TNF family members appear to contribute to hepatocellular carcinoma development. Patients with chronic hepatitis manifest elevated expression of lymphotoxin- α and - β and associated receptors. ^{64,65} In a transgenic mouse model of viral-induced hepatitis, increased levels of lymphotoxin initiated a cascade of chemokine production that culminated in a tumor-promoting inflammatory response. Blockade of this pathway with a lymphotoxin receptor antagonist inhibited the progression to carcinoma. A role for IL-1 β in tumor pathogenesis is also implicated through the association of particular polymorphisms and the risk of carcinoma development after hepatitis C infection. ⁶⁶ Finally, a contribution of adaptive immunity to disease pathogenesis was revealed through studies that showed hepatitis B–specific T cells promoted the evolution of hepatocellular carcinoma in mice transgenic for the viral envelope protein. ⁶⁷

Collectively, these investigations highlight a complex interplay of target cell damage, proinflammatory cytokines, NF- κ B activation, and immune cells in the development of liver cancer.

PROTECTIVE ANTITUMOR IMMUNE RESPONSES AND IMMUNE ESCAPE

While the tumor-promoting function for immunity is well-established, a growing body of evidence indicates that in some cases of

gastrointestinal malignancies, endogenous responses may inhibit tumor growth and perhaps modulate the clinical course of the disease. The most extensive data has been amassed in colorectal carcinomas, where intratumoral T-cell infiltrates are strongly linked with patient outcomes. Indeed, the type, density, and intratumoral location of the lymphocyte infiltrate has been shown to be a more informative biomarker than the TNM or Duke's classification. 68,69 In this context, dense infiltrates composed of cytotoxic memory CD8⁺CD45R0⁺ T cells and an associated interferon gamma-related gene signature are tightly associated with a reduced risk of recurrence after surgery and adjuvant chemotherapy, and increased overall survival. In particular, patients with early-stage cancers but an absence of T-cell infiltrates display poor outcomes, whereas subjects with significant tumor burdens but robust T-cell infiltrates manifest improved outcomes. These findings were recently confirmed in an independent cohort of patients with colon cancer, where the favorable prognostic importance of lymphocytes independent of other well-known clinicopathologic features including microsatellite instability was revealed.⁷⁰ Together, these data are consistent with the immunoediting hypothesis that was derived in model systems, and raise the possibility that tumor-specific memory T cells may be induced in some patients and perhaps contribute to disease control.

Because FoxP3⁺ regulatory T cells may restrain the antitumor activity of cytotoxic T cells, the balance of effector and suppressor cells may also prove to be a decisive factor in patient outcome. Indeed, an analysis of 308 patients with hepatocellular carcinoma showed that a high ratio of cytotoxic to regulatory T cells was associated with in-

creased survival, whereas a low ratio was linked to tumor vascular invasion, the absence of tumor encapsulation, and inferior survival.⁷¹ Moreover, a study of more than 250 histopathologically characterized pancreatic adenocarcinomas revealed that a higher density of Tregs in the primary tumor was associated with advanced tumor stage, high tumor grade, and poor survival.⁷²

In addition to eliciting a robust regulatory T-cell response, tumors may exploit other pathways to undermine cytotoxic T-cell reactions and thereby accomplish immune escape. One such mechanism is represented by structural or functional abnormalities in the HLA class I antigen processing machinery (APM). APM plays a crucial role in the synthesis and expression of the β 2-microglobulin/HLA class I heavy chain/tumor antigen-derived peptide complex. This trimolecular moiety mediates the recognition of tumor cells by cognate cytotoxic T cells. As a result of defects or lack of expression of these complexes, tumor cells may become invisible to infiltrating CD8 CD8 T cells. In addition, the frequency of APM deficiencies is increased in high-stage colorectal carcinomas and in those tumors with *Kras* mutations or microsatellite instability.

Gastrointestinal tumors may also express ligands for the stimulatory natural killer (NK) group 2, member D (NKG2D) receptor, which is expressed on NK cells and on some T cells. ^{76,77} NKG2D signaling results in perforin dependent cytolysis in NK cells and costimulation of cytotoxic T cells. The NKG2D ligands include members of the MHC class I–related chain and the UL16 binding protein families. NKG2D ligand expression is rare on healthy tissues but can be induced by infection, cellular stress, or malignant cell transformation.

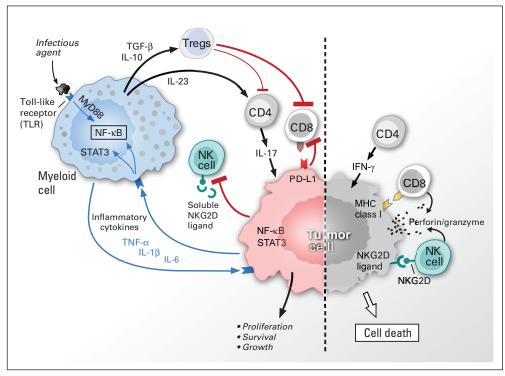


Fig 1. Dynamics of the host-tumor relationship. The overall role of immunity during cancer development reflects the balance of tumor-promoting inflammation and lymphocyte cytotoxicity. Activation of nuclear factor- κB (NF- κB) and signal transducer and activator of transcription protein 3 (STAT3) in tumor cells and myeloid elements by inflammatory cytokines and/or infection enhances cancer cell survival, proliferation, and growth. Interferon gamma (IFN- γ) –secreting CD4⁺ T cells, perforin/granzyme producing CD8⁺ T cells, and natural killer (NK) cells mediate tumor destruction. See the text for additional details. TGF, transforming growth factor; Tregs, regulatory T cells; IL, interleukin; TNF, tumor necrosis factor; PD-L1, programmed death ligand 1; NKG, natural killer group.

In human colon cancers, high MHC class I–related chain expression on primary tumors is correlated with improved overall survival. However, tumors may escape immune control in part through the shedding of NKG2D ligands, which induces downregulation of receptor expression and inhibition of NKG2D-dependent cytotoxicity. In patients with gastric cancer, the significant NKG2D downregulation on circulating CD8⁺ T was correlated with more aggressive and advanced tumors. ⁷⁹

Programmed death-1 (PD-1) is a critical negative regulator of T-cell function that is induced on lymphocyte activation. ⁸⁰ Signaling through PD-1 may be triggered through engagement of the ligands PD-L1 or PD-L2. Interestingly, some tumors upregulate the expression of PD-L1, resulting in dampened CD8⁺ T cell responses. Studies of patients with pancreatic adenocarcinoma or cholangiocarcinoma have demonstrated that high levels of intratumoral PD-L1 are associated with minimal T-cell infiltrates and poor survival. ^{81,82} Antibody blockade of PD-1/PD-L1 interactions enhances immune-mediated tumor destruction in experimental models, and initial clinical trials of this approach are underway.

THERAPEUTIC IMPLICATIONS

This review has highlighted the key roles that immunity plays in tumor promotion and tumor protection (Fig 1). One effective strategy to attenuate smoldering inflammation is prevention or eradication of persistent infections. Vaccinations against hepatitis B virus and human papilloma virus have substantively reduced the incidence of hepatocellular carcinoma and squamous cell carcinomas. A Helicobacter pylori can be eradicated with antibiotics, and if this is accomplished before irreversible alterations in the gastric mucosa, the risk of cancer can be decreased. Nonetheless, vaccines against this bacterium and other viral causes of hepatitis are urgently needed.

In certain patient populations, the use of anti-inflammatory agents can reduce the risk of developing cancer. In a randomized clinical trial, the administration of celecoxib diminished not only the cumulative adenoma incidence but also the frequency of advanced adenomas. ^{48,49} In patients with familial adenomatous polyposis, celecoxib and sulindac decrease the incidence of colorectal and duodenal

polyps, which are both precursors for invasive malignancies.^{86,87} However, a more complete understanding of the mechanisms underlying tumor-promoting inflammation has identified several compelling novel targets for intervention, including STAT-3 and NF-κB signaling pathways, and associated cytokines such as TNF family members IL-6, IL-1, IL-23, and IL-17.

Complementary strategies might focus on augmenting antitumor cytotoxic T-cell responses and inhibiting FoxP3⁺ regulatory T cells. In this context, antibody blockade of cytotoxic T lymphocyte associated—antigen 4, a key mediator of Treg suppression and a negative regulator of effector T-cell function, is in advanced stages of clinical testing in melanoma, with encouraging antitumor effects and manageable toxicities. Interestingly, inflammatory pathology in the gastrointestinal tract is an important adverse effect of this approach, which underscores from a different perspective the dual roles of immunity in cancer.

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Although all authors completed the disclosure declaration, the following author(s) indicated a financial or other interest that is relevant to the subject matter under consideration in this article. Certain relationships marked with a "U" are those for which no compensation was received; those relationships marked with a "C" were compensated. For a detailed description of the disclosure categories, or for more information about ASCO's conflict of interest policy, please refer to the Author Disclosure Declaration and the Disclosures of Potential Conflicts of Interest section in Information for Contributors.

Employment or Leadership Position: None Consultant or Advisory Role: Glenn Dranoff, Novartis (C), Schering-Plough, (C) Stock Ownership: None Honoraria: None Research Funding: None Expert Testimony: None Other Remuneration: None

AUTHOR CONTRIBUTIONS

Conception and design: Cristina Ferrone, Glenn Dranoff Collection and assembly of data: Cristina Ferrone, Glenn Dranoff Data analysis and interpretation: Cristina Ferrone, Glenn Dranoff Manuscript writing: Cristina Ferrone, Glenn Dranoff Final approval of manuscript: Cristina Ferrone, Glenn Dranoff

REFERENCES

- 1. Balkwill F, Mantovani A: Inflammation and cancer: Back to Virchow? Lancet 357:539-545, 2001
- 2. Dvorak HF: Tumors: Wounds that do not heal: Similarities between tumor stroma generation and wound healing. N Engl J Med 315:1650-1659, 1986
- 3. Ekbom A: Risk of cancer in ulcerative colitis. J Gastrointest Surg 2:312-313, 1998
- **4.** El-Omar EM, Carrington M, Chow WH, et al: The role of interleukin-1 polymorphisms in the pathogenesis of gastric cancer. Nature 412:99, 2001
- 5. Bosch FX, Ribes J, Diaz M, et al: Primary liver cancer: Worldwide incidence and trends. Gastroenterology 127:S5–S16, 2004
- **6.** Lowenfels AB, Maisonneuve P, Cavallini G, et al: Pancreatitis and the risk of pancreatic cancer: International Pancreatitis Study Group. N Engl J Med 328:1433-1437, 1993

- Lagergren J, Bergstrom R, Lindgren A, et al: Symptomatic gastroesophageal reflux as a risk factor for esophageal adenocarcinoma. N Engl J Med 340:825-831, 1999
- 8. Frisch M, Glimelius B, van den Brule AJ, et al: Sexually transmitted infection as a cause of anal cancer. N Engl J Med 337:1350-1358, 1997
- **9.** Bergquist A, Ekbom A, Olsson R, et al: Hepatic and extrahepatic malignancies in primary sclerosing cholangitis. J Hepatol 36:321-327, 2002
- **10.** Koebel CM, Vermi W, Swann JB, et al: Adaptive immunity maintains occult cancer in an equilibrium state. Nature 450:903-907, 2007
- 11. Dougan M, Dranoff G: Immune therapy for cancer. Annu Rev Immunol 27:83-117, 2009
- **12.** Tracey KJ, Beutler B, Lowry SF, et al: Shock and tissue injury induced by recombinant human cachectin. Science 234:470-474, 1986
- **13.** Okayasu I, Hatakeyama S, Yamada M, et al: A novel method in the induction of reliable experimental acute and chronic ulcerative colitis in mice. Gastroenterology 98:694-702, 1990

- **14.** Boivin GP, Washington K, Yang K, et al: Pathology of mouse models of intestinal cancer: Consensus report and recommendations. Gastroenterology 124: 762-777, 2003
- **15.** Popivanova BK, Kitamura K, Wu Y, et al: Blocking TNF-alpha in mice reduces colorectal carcinogenesis associated with chronic colitis. J Clin Invest 118:560-570, 2008
- **16.** Mantovani A, Allavena P, Sica A, et al: Cancer-related inflammation. Nature 454:436-444,
- **17.** Ghosh S, Karin M: Missing pieces in the NF-kappaB puzzle. Cell 109:S81-S96, 2002 (suppl)
- **18.** Greten FR, Eckmann L, Greten TF, et al: IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. Cell 118: 285-296, 2004
- 19. Eaden J, Abrams K, Ekbom A, et al: Colorectal cancer prevention in ulcerative colitis: A case-control study. Aliment Pharmacol Ther 14: 145-153, 2000

- **20.** Kopp E, Ghosh S: Inhibition of NF-kappa B by sodium salicylate and aspirin. Science 265:956-959, 1994
- 21. Jacobs EJ, Thun MJ, Bain EB, et al: A large cohort study of long-term daily use of adult-strength aspirin and cancer incidence. J Natl Cancer Inst 99:608-615, 2007
- **22.** Kim S, Takahashi H, Lin WW, et al: Carcinoma-produced factors activate myeloid cells through TLR2 to stimulate metastasis. Nature 457: 102-106, 2009
- 23. Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, et al: Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. Cell 118:229-241, 2004
- 24. Rakoff-Nahoum S, Medzhitov R: Regulation of spontaneous intestinal tumorigenesis through the adaptor protein MyD88. Science 317:124-127, 2007
- **25.** Pull SL, Doherty JM, Mills JC, et al: Activated macrophages are an adaptive element of the colonic epithelial progenitor niche necessary for regenerative responses to injury. Proc Natl Acad Sci U S A 102:99-104, 2005
- **26.** Fukata M, Michelsen KS, Eri R, et al: Toll-like receptor-4 is required for intestinal response to epithelial injury and limiting bacterial translocation in a murine model of acute colitis. Am J Physiol Gastrointest Liver Physiol 288:G1055–G1065, 2005
- 27. Yu H, Pardoll D, Jove R: STATs in cancer inflammation and immunity: A leading role for STAT3. Nat Rev Cancer 9:798-809, 2009
- 28. Catlett-Falcone R, Landowski TH, Oshiro MM, et al: Constitutive activation of Stat3 signaling confers resistance to apoptosis in human U266 myeloma cells. Immunity 10:105-115, 1999
- **29.** Kujawski M, Kortylewski M, Lee H, et al: Stat3 mediates myeloid cell-dependent tumor angiogenesis in mice. J Clin Invest 118:3367-3377, 2008
- **30.** Grivennikov S, Karin E, Terzic J, et al: IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. Cancer Cell 15:103-113, 2009
- **31.** Wu S, Rhee KJ, Albesiano E, et al: A human colonic commensal promotes colon tumorigenesis via activation of T helper type 17 T cell responses. Nat Med 15:1016-1022, 2009
- **32.** Grady WM, Myeroff LL, Swinler SE, et al: Mutational inactivation of transforming growth factor beta receptor type II in microsatellite stable colon cancers. Cancer Res 59:320-324, 1999
- **33.** Chung YC, Chang YF: Serum interleukin-6 levels reflect the disease status of colorectal cancer. J Surg Oncol 83:222-226, 2003
- **34.** Galizia G, Orditura M, Romano C, et al: Prognostic significance of circulating IL-10 and IL-6 serum levels in colon cancer patients undergoing surgery. Clin Immunol 102:169-178, 2002
- **35.** Langowski JL, Zhang X, Wu L, et al: IL-23 promotes tumour incidence and growth. Nature 442:461-465, 2006
- **36.** Chen Z, O'Shea JJ: Th17 cells: A new fate for differentiating helper T cells. Immunol Res 41:87-102, 2008
- **37.** Wang L, Yi T, Kortylewski M, et al: IL-17 can promote tumor growth through an IL-6-Stat3 signaling pathway. J Exp Med 206:1457-1464, 2009
- **38.** Kortylewski M, Xin H, Kujawski M, et al: Regulation of the IL-23 and IL-12 balance by Stat3 signaling in the tumor microenvironment. Cancer Cell 15:114-123, 2009
- **39.** Glocker EO, Kotlarz D, Boztug K, et al: Inflammatory bowel disease and mutations affecting the interleukin-10 receptor. N Engl J Med 361:2033-2045, 2009

- **40.** Berg DJ, Kuhn R, Rajewsky K, et al: Interleukin-10 is a central regulator of the response to LPS in murine models of endotoxic shock and the Shwartzman reaction but not endotoxin tolerance. J Clin Invest 96:2339-2347, 1995
- **41.** Kuhn R, Lohler J, Rennick D, et al: Interleukin-10-deficient mice develop chronic enterocolitis. Cell 75:263-274, 1993
- **42.** Spencer SD, Di Marco F, Hooley J, et al: The orphan receptor CRF2-4 is an essential subunit of the interleukin 10 receptor. J Exp Med 187:571-578, 1998
- **43.** Moore KW, de Waal Malefyt R, Coffman RL, et al: Interleukin-10 and the interleukin-10 receptor. Annu Rev Immunol 19:683-765, 2001
- **44.** Fiorentino DF, Zlotnik A, Vieira P, et al: IL-10 acts on the antigen-presenting cell to inhibit cytokine production by Th1 cells. J Immunol 146:3444-3451, 1991
- **45.** Becker C, Fantini MC, Schramm C, et al: TGF-beta suppresses tumor progression in colon cancer by inhibition of IL-6 trans-signaling. Immunity 21:491-501, 2004
- **46.** Gounaris E, Blatner NR, Dennis K, et al: T-regulatory cells shift from a protective anti-inflammatory to a cancer-promoting proinflammatory phenotype in polyposis. Cancer Res 69:5490-5497, 2009
- **47.** Erdman SE, Sohn JJ, Rao VP, et al: CD4+CD25+ regulatory lymphocytes induce regression of intestinal tumors in ApcMin/+ mice. Cancer Res 65:3998-4004, 2005
- **48.** Bertagnolli MM, Eagle CJ, Zauber AG, et al: Five-year efficacy and safety analysis of the Adenoma Prevention with Celecoxib Trial. Cancer Prev Res (Phila Pa) 2:310-321, 2009
- **49.** Bertagnolli MM, Eagle CJ, Zauber AG, et al: Celecoxib for the prevention of sporadic colorectal adenomas. N Engl J Med 355:873-884, 2006
- **50.** Maltby S, Khazaie K, McNagny KM: Mast cells in tumor growth: Angiogenesis, tissue remodelling and immune-modulation. Biochim Biophys Acta 1796:19-26, 2009
- **51.** Fox JG, Wang TC: Inflammation, atrophy, and gastric cancer. J Clin Invest 117:60-69, 2007
- **52.** Fox JG, Wang TC: Helicobacter pylori: Not a good bug after all! N Engl J Med 345:829-832, 2001
- **53.** Figueiredo C, Machado JC, Pharoah P, et al: Helicobacter pylori and interleukin 1 genotyping: An opportunity to identify high-risk individuals for gastric carcinoma. J Natl Cancer Inst 94:1680-1687, 2002
- **54.** Dinarello CA: Biologic basis for interleukin-1 in disease. Blood 87:2095-2147, 1996
- **55.** Dolcetti L, Marigo I, Mantelli B, et al: Myeloid-derived suppressor cell role in tumor-related inflammation. Cancer Lett 267:216-225, 2008
- **56.** Tu S, Bhagat G, Cui G, et al: Overexpression of interleukin-1beta induces gastric inflammation and cancer and mobilizes myeloid-derived suppressor cells in mice. Cancer Cell 14:408-419, 2008
- **57.** Bronte-Tinkew DM, Terebiznik M, Franco A, et al: Helicobacter pylori cytotoxin-associated gene A activates the signal transducer and activator of transcription 3 pathway in vitro and in vivo. Cancer Res 69:632-639, 2009
- **58.** Sawai N, Kita M, Kodama T, et al: Role of gamma interferon in Helicobacter pylori-induced gastric inflammatory responses in a mouse model. Infect Immun 67:279-285. 1999
- **59.** Smythies LE, Waites KB, Lindsey JR, et al: Helicobacter pylori-induced mucosal inflammation is Th1 mediated and exacerbated in IL-4, but not

- IFN-gamma, gene-deficient mice. J Immunol 165: 1022-1029, 2000
- **60.** Berg DJ, Lynch NA, Lynch RG, et al: Rapid development of severe hyperplastic gastritis with gastric epithelial dedifferentiation in Helicobacter felis-infected IL-10(-/-) mice. Am J Pathol 152:1377-1386, 1998
- **61.** Eaton KA, Mefford M, Thevenot T: The role of T cell subsets and cytokines in the pathogenesis of Helicobacter pylori gastritis in mice. J Immunol 166:7456-7461, 2001
- **62.** Pikarsky E, Porat RM, Stein I, et al: NF-kappaB functions as a tumour promoter in inflammation-associated cancer. Nature 431:461-466, 2004
- **63.** Maeda S, Kamata H, Luo JL, et al: IKKbeta couples hepatocyte death to cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. Cell 121:977-990, 2005
- **64.** Haybaeck J, Zeller N, Wolf MJ, et al: A lymphotoxin-driven pathway to hepatocellular carcinoma. Cancer Cell 16:295-308, 2009
- **65.** Fu YX, Huang G, Wang Y, et al: B lymphocytes induce the formation of follicular dendritic cell clusters in a lymphotoxin alpha-dependent fashion. J Exp Med 187:1009-1018, 1998
- **66.** Wang Y, Kato N, Hoshida Y, et al: Interleukin-1beta gene polymorphisms associated with hepatocellular carcinoma in hepatitis C virus infection. Hepatology 37:65-71, 2003
- **67.** Nakamoto Y, Guidotti LG, Kuhlen CV, et al: Immune pathogenesis of hepatocellular carcinoma. J Exp Med 188:341-350, 1998
- **68.** Pages F, Berger A, Camus M, et al: Effector memory T cells, early metastasis, and survival in colorectal cancer. N Engl J Med 353:2654-2666, 2005
- **69.** Galon J, Costes A, Sanchez-Cabo F, et al: Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. Science 313:1960-1964, 2006
- **70.** Ogino S, Nosho K, Irahara N, et al: Lymphocytic reaction to colorectal cancer is associated with longer survival, independent of lymph node count, microsatellite instability, and CpG island methylator phenotype. Clin Cancer Res 15:6412-6420, 2009
- **71.** Gao Q, Qiu SJ, Fan J, et al: Intratumoral balance of regulatory and cytotoxic T cells is associated with prognosis of hepatocellular carcinoma after resection. J Clin Oncol 25:2586-2593, 2007
- **72.** Hiraoka N, Onozato K, Kosuge T, et al: Prevalence of FOXP3+ regulatory T cells increases during the progression of pancreatic ductal adenocarcinoma and its premalignant lesions. Clin Cancer Res 12:5423-5434, 2006
- 73. Salter RD, Howell DN, Cresswell P: Genes regulating HLA class I antigen expression in T-B lymphoblast hybrids. Immunogenetics 21:235-246, 1985
- **74.** Sadasivan B, Lehner PJ, Ortmann B, et al: Roles for calreticulin and a novel glycoprotein, tapasin, in the interaction of MHC class I molecules with TAP. Immunity 5:103-114, 1996
- **75.** Kloor M, Becker C, Benner A, et al: Immunoselective pressure and human leukocyte antigen class I antigen machinery defects in microsatellite unstable colorectal cancers. Cancer Res 65:6418-6424, 2005
- **76.** Bauer S, Groh V, Wu J, et al: Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. Science 285:727-729, 1999
- 77. Raulet DH: Roles of the NKG2D immunoreceptor and its ligands. Nat Rev Immunol 3:781-790, 2003

Immunity in Cancer Pathogenesis

- **78.** McGilvray RW, Eagle RA, Watson NF, et al: NKG2D ligand expression in human colorectal cancer reveals associations with prognosis and evidence for immunoediting. Clin Cancer Res 15:6993-7002, 2009
- **79.** Osaki T, Saito H, Yoshikawa T, et al: Decreased NKG2D expression on CD8+ T cell is involved in immune evasion in patients with gastric cancer. Clin Cancer Res 13:382-387, 2007
- **80.** Greenwald RJ, Freeman GJ, Sharpe AH: The B7 family revisited. Annu Rev Immunol 23:515-548, 2005
- **81.** Nomi T, Sho M, Akahori T, et al: Clinical significance and therapeutic potential of the programmed death-1 ligand/programmed death-1 path-

- way in human pancreatic cancer. Clin Cancer Res 13:2151-2157, 2007
- **82.** Ye Y, Zhou L, Xie X, et al: Interaction of B7–H1 on intrahepatic cholangiocarcinoma cells with PD-1 on tumor-infiltrating T cells as a mechanism of immune evasion. J Surg Oncol 100:500-504, 2009
- **83.** Chang MH, You SL, Chen CJ, et al: Decreased incidence of hepatocellular carcinoma in hepatitis B vaccinees: A 20-year follow-up study. J Natl Cancer Inst 101:1348-1355, 2009
- **84.** Chang Y, Brewer NT, Rinas AC, et al: Evaluating the impact of human papillomavirus vaccines. Vaccine 27:4355-4362, 2009
- **85.** Roggero E, Zucca E, Pinotti G, et al: Eradication of Helicobacter pylori infection in primary low-

- grade gastric lymphoma of mucosa-associated lymphoid tissue. Ann Intern Med 122:767-769, 1995
- **86.** Phillips RK, Wallace MH, Lynch PM, et al: A randomised, double blind, placebo controlled study of celecoxib, a selective cyclooxygenase 2 inhibitor, on duodenal polyposis in familial adenomatous polyposis. Gut 50:857-860, 2002
- **87.** Debinski HS, Trojan J, Nugent KP, et al: Effect of sulindac on small polyps in familial adenomatous polyposis. Lancet 345:855-856, 1995
- 88. Hodi FS, Dranoff G: Combinatorial cancer immunotherapy. Adv Immunol 90:337-360, 2006
- 89. Korman A, Peggs K, Allison JP: Checkpoint blockade in cancer immunotherapy. Adv Immunol 90:293-335. 2006

Sign up for Alerts About Your Topic of Interest

Learn about new research in your field as it becomes available. Subscribe to a *JCO* e-mail alert to be notified immediately when new articles within your area of interest are posted.

Receive notification when:

- > JCO releases a new issue's Table of Contents.
- A new issue of JCO is posted online.
- New articles are published online ahead of print publication.
- New content in your subspecialty is published.
- An article is published online from an author of interest.

Go to jco.org/alerts to sign up.

ASCO®

American Society of Clinical Oncology